# Sepsis in Infants: Analysis of Bacterial Pathogens and their Antibiotic Susceptibility, A Study at Government Tertiary Care Hospital, Karachi

Masood Hassan Rao,<sup>1</sup> Sadia Khan,<sup>2</sup> Tooba Waseem,<sup>2</sup> Sabah Naeem<sup>2</sup> and Sadaf Sabir<sup>2</sup>

### ABSTRACT

**Objective:** To assess the frequency of causative bacterial pathogen of sepsis in infant, their antibiotic susceptibility and to determine resistance pattern in commonly used antibiotics.

Place and Duration of Study: Central Lab Civil Hospital, Karachi, 6 months.

Study Design: Retrospective Descriptive observational study.

**Patients and Method:** All 1414 reports of blood samples send for culture/sensitivity of infants admitted in Civil Hospital Karachi during the study period were scrutinized for bacterial pathogen, their frequency, antibiotic susceptibility and resistance pattern.

**Result:** Out of 1414 infants, 604 (42.7%) had positive blood culture. Gram positive bacteria were predominant (54.1%) than gram negative (45.9%). Male: female ratio was found to be approximately 1:0.9. Total 9 organisms were isolated, in which staphylococcus aureus predominates followed by Pseudomonas aeruginosa and Escherichia coli respectively. The overall sensitivity of the organism to Amikacin and Cefotaxime were 60.87% and 36.67% respectively which are currently in use as empirical therapy in pediatric ward of CHK. The organisms were most sensitive to Vancomycin (95.54%), Sparfloxacin (94.16%), Linezolid (93.56%), while mostly resistant to kanamycin (56.21%), cephalosporins (55.9%), Gentamycin (54.31%) and amoxicillin (51.11%).

**Conclusion:** Gram positive organisms were identified as the major threat for sepsis in infants. An emerging pattern of resistance was observed against commonly used antibiotic so there is a need to control the spread of these resistant strains through infection control programs and continuous monitoring of drug resistant patterns.

Key words: Septicemia in infants, culture and sensitivity pattern, drug resistance.

*How to cite this article:* Rao MH, Khan S, Waseem T, Naeem S, Sabir S. Sepsis in Infants: Assessment of Bacterial Pathogens and their Antibiotic Susceptibility, A Study at Government Tertiary Care Hospital, Karachi J Dow Uni Health Sci 2013; 7(1): 35-40.

### **INTRODUCTION**

Sepsis is defined as systemic inflammatory response syndrome resulting from suspected or proven infection.<sup>1</sup> According to an estimate of World Health Organization (WHO), out of 4 million neonatal deaths occurring around the world every year,<sup>2</sup> approximately 98% of these deaths occur in developing countries.<sup>3</sup> Neonatal sepsis is a predominant cause of morbidity and mortality and reason for frequent hospital admissions of children in developing countries.<sup>4</sup>

**Correspondence:** Sadia Khan, Final Year MBBS Student, Dow University of Health Sciences, Karachi, Pakistan.

E-mail: dr.sadiakhan@hotmail.com

Pakistan accounts for about 7% of global neonatal deaths.<sup>5</sup> One third of these deaths occur due to infections.<sup>6</sup> Infants are at highest risk, with 10 times higher than that of older children. Low and very low-birth-weight (VLBW) babies make up nearly one fourth of the pediatric sepsis population.<sup>7</sup>

Sepsis in children is a life-threatening emergency and any delay in treatment may cause death. Initial signs of sepsis are slight and nonspecific. Therefore, in suspected cases, empirical antibiotic therapy should begin immediately without awaiting the results<sup>8</sup> of blood culture and sensitivity reports. Early treatment and appropriate use of antibiotics minimize the risk of severe morbidity and mortality in sepsis and reduce the emergence of multi-drug resistant organisms in intensive care units by rational antibiotic use.<sup>9</sup>

The distribution of pathogens causing sepsis in a specific hospital unit is usually considered when empiric antibiotics are selected.<sup>10</sup> The bacteriological profile

Journal of the Dow University of Health Sciences Karachi 2013, Vol. 7 (1): 35-40

<sup>1</sup> Principal Research Officer, PMRC Research Center, Dow Medical College, Karachi, Pakistan.

<sup>2</sup> Final Year Students, Dow University of Health Sciences, Karachi, Pakistan.

of neonatal septicemia is constantly under change with advances in early diagnosis and treatment. Thus the rational protocol for sepsis management must be based on adequate knowledge of the causative organism and their antibiotic sensitivity pattern in related area.<sup>4</sup>

In Pakistan, a very little data is available on infant's sepsis and their antibiotic susceptibility and resistance. So we have planned to provide the health management an open access to those antibiotics which are more sensitive to common bacteria now a day.

# **METHODOLOGY**

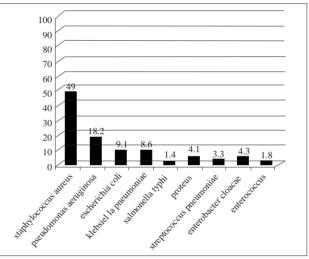
The study was conducted at central lab CHK during the period of December 2010 - May 2011. A 5ml Blood sample of infants under one year of age admitted in pediatric wards of CHK with clinical sign and symptoms of sepsis as defined by WHO, were received by the lab for Culture and sensitivity. The blood was aseptically drawn and inoculated into the BD BACTEC culture bottle and incubated for 5days. Sub culture was done on Blood Agar, Chocolate Agar and Mc.Conkeys Agar and incubated at 37<sup>0</sup>C aerobically. The plates were then examined for any bacterial growth. Gram staining and various biochemical and serological test were done for identification and antibiotic susceptibility was checked by Kirby Baur Disc diffusion technique. Blood culture reports were developed and a copy of the same was sent to the concerned unit. The data was analyzed through SPSS version 16.0 and frequency and percentages were calculated for each qualitative variable. The effective antibiotics were determined by applying chi-square test with significant values P < 0.05. Isolates showing intermediate level of resistance were classified as sensitive.

# RESULTS

During the study period of 6 months, total 1414 blood samples of infants (under one year of age) suspected with sepsis were included. Out of these, 604 (42.7%) had positive blood culture. Of these, 322 (53.3%) were males and 282 (46.7%) were females. Male: female ratio was approximately 1:0.9. In 604 positive blood culture reports, Gram positive bacteria was found in 54.1% as compared to gram negative (45.9%). A total 9 organisms were isolated; in which staphylococcus aureus (49%) predominates followed by Pseudomonas aeruginosa (18.2%) and Escherichia coli (9.1%) respectively (Graph 1).

The antibiotic sensitivity pattern of the bacterial isolates was also analyzed. The overall sensitivity of the organisms was found with Amikacin and Cefotaxime (60.87% and 36.67% respectively) which are currently in use as empirical therapy in pediatric ward of CHK. Overall, the organisms were most sensitive to Vancomycin (95.54%), Sparfloxacin (94.16%), Linezolid (93.56%), while resistance was mostly to cephalosporins, (Cefaclor 81.55%, Cephradine 78.85%, Cefixime 75.69%, and Cefuroxime 75.47%), kanamycin 56.21%, Gentamycin 54.31% and amoxicillin 51.11%. (Graph 2)

According to the Sensitivity and resistance pattern of bacterial isolates, sparfloxacin and vancomycin were found to be most sensitive with most of the bacterias, where as amoxicillin and Kanamycin was the most resistant to the isolated bacteria. (Table I)



Graph 1: Frequency of Bacterial Pathogens (In Percentages)

# **DISCUSSION**

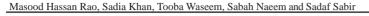
Sepsis is a predominant cause of morbidity and mortality and reason for frequent hospital admissions of children. It is a life-threatening emergency and any delay in treatment may cause death. In our study positive blood cultures were found to be 42.7%. This suggests that sepsis in infants remained major cause of morbidity in hospitals. Similar studies were conducted in 2002 and 2008 in CHK which showed 55.2%<sup>11</sup> and 30.9%<sup>12</sup> positive cultures respectively, showing these percentages are not varying much in the same hospital within 10 years. Different researches in Pakistan showed different results such as 40% positive cultures in Islamabad,<sup>13</sup> 32% in Lahore,<sup>14</sup> 62.8% in Peshawar.<sup>15</sup>

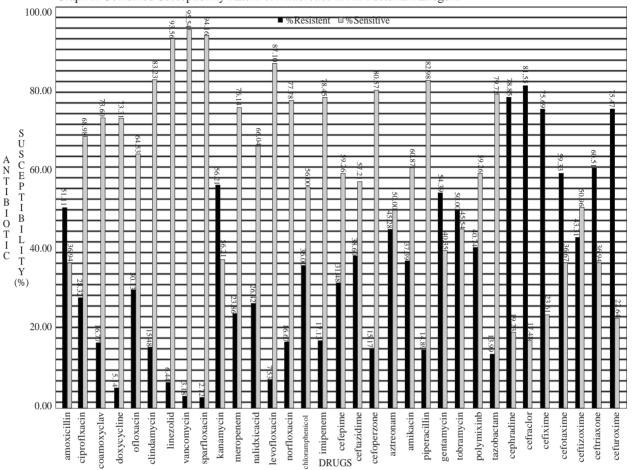
In our study, all cultures yielded single pathogen. This is similar to other studies conducted in Karachi^{12} and Lahore.  $^{16}$ 

Gram positive cocci were most frequent 54.1% which is in contrast to previous studies in CHK,<sup>11,12</sup> Pakistan<sup>14,15,17</sup> and neighboring countries.<sup>18-20</sup> However, in a study<sup>21</sup> gram positive organisms were found to be the main cause of neonatal sepsis in Karachi. Similar predominance of Gram-positive organisms was reported in different centers in Nigeria.<sup>22,23</sup>

|    | Bacteria                 | Most Sensitive Antibiotic | Total Used | Sensitivity | Most Resistant Antibiotic | Total used | Resistance |
|----|--------------------------|---------------------------|------------|-------------|---------------------------|------------|------------|
| 1. | Staphylococcus Aureus    | 1.sparfloxacin            | 116        | 97.41%      | 1.kanamycin               | 270        | 57.78%     |
|    | 1 5                      | 2.vancomycin              | 292        | 97.26%      | 2.amoxicillin             | 294        | 52.04%     |
|    |                          | 3.linezolid               | 293        | 93.52%      | 3.ciprofloxacin           | 264        | 39.05%     |
|    |                          | 4.clindamycin             | 288        | 85.42%      | 4.ofloxacin               | 291        | 38.14%     |
|    |                          | 5.coamoxyclav             | 290        | 83.45%      | 5.cefepime                | 66         | 34.85%     |
| 2. | Pseudomonas Aeruginosa   | 1.sparfloxacin            | 47         | 93.62%      | 1.Gentamycin              | 110        | 54.55%     |
|    |                          | 2.ceftazidime             | 106        | 77.36%      | 2.tobramycin              | 108        | 49.07%     |
|    |                          | 3.ciprofloxacin           | 93         | 90.32%      | 3.aztreonam               | 103        | 44.66%     |
|    |                          | 4.Pipera/tazobactam       | 58         | 89.66%      | 4.polymixinb              | 106        | 41.51%     |
|    |                          | 5.amikacin                | 110        | 64.55%      | 5.imipenem                | 108        | 18.52%     |
| 3. | Escherichia coli         | 1.sparfloxacin            | 26         | 96.15%      | 1.cefixime                | 53         | 77.36%     |
|    |                          | 2.levofloxacin            | 51         | 90.20%      | 2.cefaclor                | 53         | 71.70%     |
|    |                          | 3.cefoperzone             | 35         | 77.14%      | 3.cephradine              | 52         | 67.31%     |
|    |                          | 4.ofloxacin               | 53         | 75.47%      | 4.cefuroxime              | 51         | 62.75%     |
|    |                          | 5.Pipera/tazobactam       | 27         | 74.07%      | 5.ceftriaxone             | 51         | 52.94%     |
| 4. | Klebsiella pneumoniae    | 1.levofloxacin            | 48         | 91.67%      | 1.cefaclor                | 48         | 91.67%     |
|    |                          | 2.ofloxacin               | 49         | 85.71%      | 2.cefixime                | 48         | 93.75%     |
|    |                          | 3.ciprofloxacin           | 45         | 82.22%      | 3.cefuroxime              | 49         | 98.14%     |
|    |                          | 4.cefoperzone             | 13         | 76.92%      | 4.ceftriaxone             | 48         | 87.54%     |
|    |                          | 5.ceftizoxime             | 46         | 50%         | 5.cefotaxime              | 45         | 80%        |
| 5. | Salmonella typhi         | 1.ofloxacin               | 7          | 100%        | 1. amoxicillin            | 9          | 66.67%     |
| 5. | Sumonena typin           | 2.sparfloxacin            | 5          | 100%        | 2.ceftizoxime             | 8          | 50%        |
|    |                          | 3. imipenem               | 9          | 88.9%       | 3.ceftizoxime             | 2          | 50%        |
|    |                          | 4. levofloxacin           | 9          | 88.9%       | 4.chloramphenicol         | 9          | 44.4%      |
|    |                          | 5. norfloxacin            | 9          | 88.9%       | 5.cefixime                | 9          | 44.4%      |
| 6. | Proteus                  | 1.cefoperzone             | 20         | 95%         | 1.amoxicillin             | 15         | 60%        |
| 0. | Floteus                  | 2.norfloxacin             |            |             | 2.cefotaxime              |            |            |
|    |                          |                           | 18         | 94.44%      | 3.ceftriaxone             | 19         | 57.89%     |
|    |                          | 3.Pipera/tazobactam       |            | 94.44%      |                           | 25         | 56%        |
|    |                          | 4.levofloxacin            | 25         | 92%         | 4.ceftizoxime             | 24         | 45.83%     |
| 7  | <u> </u>                 | 5.ofloxacin               | 24         | 87.5%       | 5.cefixime                | 11         | 45.45%     |
| 7. | Streptococcus pneumoniae | 1.vancomycin              | 20         | 95%         | 1.kanamycin               | 18         | 38.89%     |
|    |                          | 2.linezolid               | 20         | 95%         | 2.ofloxacin               | 20         | 20%        |
|    |                          | 3.coamoxyclav             | 20         | 90%         | 3.clindamycin             | 20         | 25%        |
|    |                          | 4.amoxicillin             | 18         | 88.89%      | 4                         |            |            |
| 0  |                          | 5.ciprofloxacin           | 20         | 75%         | 5                         |            | ~~~~       |
| 8. | Enterobacter cloacae     | 1.levofloxacin            | 26         | 96.15%      | 1.amoxicillin             | 23         | 60.8%      |
|    |                          | 2.imipenem                | 26         | 92.30%      | 2.cefixime                | 23         | 60.86%     |
|    |                          | 3.cefoperzone             | 25         | 92%         | 3.cefotaxime              | 26         | 50%        |
|    |                          | 4.Piper/tazobactam        | 6          | 100%        | 4.ceftizoxime             | 26         | 46.15%     |
|    |                          | 5.ofloxacin               | 22         | 86.36%      | 5.ceftriaxone             | 25         | 40%        |
| 9. | Enterococcus             | 1.imipenem                | 11         | 81.81%      | 1 -                       |            |            |
|    |                          | 2.Coamoxyclav             | 11         | 72.72%      | 2                         |            |            |
|    |                          | 2.Coamoxyclav             | 11         | 72.72%      | 2<br>3                    |            |            |

Table I: Sensitivity Pattern of Bacterial Isolates





Graph 2: Combined Susceptibility Pattern of Antibiotics in All Bacterial Pathogens

The male gender Preponderance in our study is similar with few other previous studies.<sup>13,15,16,18-20</sup>

Staphylococcus aureus emerged as most frequent causative agent of septicemia in our study (49%). As it is a normal flora of skin, lack of awareness about safety protocols during medical procedures in overall society may be a cause of such high prevalence. Other two common organisms were Pseudomonas auregnosa (18.2%) and Escherichia coli (9.1%). We found change in frequency of different organism in CHK with time as in 2008, Escheria coli and Staphylococcus aureus were most common<sup>12</sup> while in 2005, klebsiella pneumoniae was the major isolate (24%) followed by staphylococcus aureus (22%).<sup>11</sup> Studies outside Karachi showed high incidence of Escherichia coli in Lahore<sup>14</sup> and Multan<sup>9</sup> and enterobacter in Islamabad.<sup>13</sup> Among neighboring countries, enterobacter was most common in Iran,<sup>19</sup> pseudomonas and klebsiella in India<sup>18</sup> and Bangladesh.<sup>20</sup> Thus different geographical areas have different prevalence.

Group B streptococcus (GBS) was not isolated from any culture in our study, the same has been reported in most of the studies in Pakistan.<sup>14,11,15</sup> However, a single isolate was reported in a previous study at CHK,<sup>12</sup> AKU<sup>24</sup> Karachi and Peshawar<sup>25</sup> which showed incidence of sepsis with GBS. In contrast, western countries reported GBS as their frequent isolate.<sup>26</sup> This difference may be because of presence of strains of low virulence, low prevalence of colonization of GBS in pregnant women or simply because of low socioeconomic condition resulting in death of most early onset sepsis (EOS) babies at home which remain unreported.

In our study, Staphylococcus aureus showed high resistance to ampicillin (75%) which corresponds with other studies.<sup>11,13,19,20</sup> This demonstrates that the use of this drug is now inappropriate. However, it remained sensitive to vancomycin (97%) as reported by others also.<sup>10,12,14,17</sup> This suggests that guidelines should be made for prudent use of Vancomycin, in an attempt to prevent the spread of vancomycin resistant strains.

Over all, Flouroquinolones remained highly sensitive in most of the organisms.<sup>13,25</sup> Among them, ciprofloxacin showed overall 68.9% sensitivity. This indicates that it may be a good choice for septicemia in infants as it is inexpensive, penetrates CSF, has good oral bioavailability<sup>27</sup> and has an acceptable safety profile in infants.<sup>28,29</sup>

There is considerable degree of resistance in Cephalosporins ranging from 15% (cefoperzone) to 81.55% (cefaclor). Similar degree of resistance is reported in different studies as well.<sup>9,13,15,17</sup> Cefotaxime (an empirical therapy drug in CHK) showed considerable resistance (59.33%) against gram negative rods. This emerging pattern of resistance is supported by the work of A. Mahmood et al,<sup>17</sup> Batool A et al<sup>11</sup> and others<sup>9,13,15,19</sup> suggesting that the cautious use of cefotaxime should be promoted as resistance to this may evoke a great challenge for physicians in choosing antimicrobial therapy, as the treatment options for cephalosporin resistant strains are limited.

Aminoglycosides showed moderate degree of resistance against most of gram negative rods ranging from 37% (amikacin) to 56% (kanamycin). In contrast, in a study<sup>11</sup> conducted in in CHK reported significant resistance for aminoglycosides, same was found in other studies.<sup>14,15,30</sup> However, in our study among aminoglycosides, Amikacin showed good sensitivity and this pattern is consistent to that found by Mahmood et al in Karachi<sup>17</sup> and Shams et al<sup>13</sup> in Islamabad.

Amikacin and Cefotaxime are currently used as empirical treatment in infants at our hospital which showed 60.87% and 36.67% sensitivity respectively. However, the observation in the present study may not be conclusive because the sensitivity of Amikacin and cefotaxime is not tested for gram positive cocci (Staph. Aureus) in CHK Lab, hence, it is highly recommended that lab should check these drugs, as they are important therapy against Staphylococcus and pseudomonas and we are lacking current susceptibility pattern of most prevalent organisms to them, in our setup.

### **CONCLUSION**

We conclude from our study that Staphylococcus aureus is a major cause of sepsis in infants admitted in CHK. Moreover, the frequency of bacterial pathogens and antibiotic susceptibility is varying geographically and with time. Also, the antibiotic susceptibility profile of different organisms is showing considerable resistance to conventional antibiotics. Therefore, routine bacterial surveillance and study of their resistance patterns is essential for formation of antibiotic policy guidelines, and such a study should be carried out in every hospital on periodic basis. In this context, we propose the continuation of use of Amikacin in our current empirical therapy. In order to reduce morbidity and mortality it is crucial to consider latest guidelines and prudent use of antibiotics focused on accurate dosage.

# LIMITATIONS

According to our limitations, we were unable to differentiate between early and late onset of sepsis as records were available for age in years only. Secondly, in our setup, methicillin is not tested for staphylococci so MRSA strains prevalence is still indefinable.

#### REFERENCES

- 1 Enrione MA, Powell KR, Sepsis, Septic shock and Systemic inflammatory response syndrome: Nelson Text book of pediatrics. volume 1, 18th edition; Pg. NO.1094.
- 2 World Health Organization. Perinatal mortality. A listing of available information. Geneva, WHO 1996:32-6. (WHO/ FRH/MSM/96.7)
- 3 Black RE, Moris SS, Bryce J. Where and why are 10 million children dying every year? Lancet 2003; 361: 2226-34.
- 4 Agrawal R, Sarkar N, Deorary AK, Pau VK, Sepsis in newborn: Ind J Pediatric, 2000; 68:1143-7.
- 5 Jehan I, harris H, Salat S, Zeb A, Mobeen N, Pasha O, et al. Neonatal mortality, risk factors and causes: a prospective population-based cohort study in urban Pakistan: Bull World Health Organ 2009; 87:130-8.
- 6 Jalil F, Lindblad BS, Hanson LA, Khan SR, Yaqoob M, Karlberg J. Early child health in Lahore, Pakistan: IX Perinatal events. Acta Paediatrica 1993; 82:95-107.
- 7 Watson, Scott R, Carcillo, Joseph A. Scope and epidemiology of pediatric sepsis: Pediatric Critical Care Medicine 2005; 69:3-5.
- 8 Yurdakök M. Antibiotic use in neonatal sepsis: Turk J Pediatr 1998; 40:17-33.
- 9 Sheikh AM, Javed T, Afzal MF, Sheikh CA, Course and Complications of Early Onset Neonatal Sepsis: A Descriptive Study, Annals 2010; 16:307-10.
- 10 Gary KM, Stephen BE, Surka AE. Fulminant Late-Onset Sepsis in a Neonatal Intensive Care Unit, 1988-1997, and the Impact of Avoiding Empiric Vancomycin Therapy: Pediatrics 2000; 106;1387.
- 11 Batool A, Akram DS, Arif F. Changing antibiotic sensitivity of organisms causing neonatal sepsis: A hospital based study. Pak Paed J 2005; 29:57-61.
- 12 Abdullah FE, Taj Y, Sharafat S. Current pattern of bloodstream infections in a tertiary care hospital of Karachi and clinical significance of positive blood cultures: J Dow Univ Health Sci 2010; 4:25-30.
- 13 Shams R, Khan N, Hussain S: Bacteriology & Anti-Microbial Susceptibility of Neonetal Septicemia in NICU, PIMS, Islamabad-A Tertiary Care Hospital of Pakistan: Ann Pak Inst Med Sci 2010; 6:191-5.
- Ahmad A, Hussain W, Lamichhane A, Aslam M, Riaz L. Use of Antibiotics in Neonatal Sepsis at Neonatal Unit of A Tertiary Care Hospital: Pak pead J 2011; 35:3-7.

- 15 Rahman S, Hameed A, Roghani MT, et al. Multidrug resistant neonatal sepsis in Peshawar, Pakistan: Arch Dis Child Fetal Neonatal Ed 2002; 87:52-4.
- 16 Chaudhry I, Chaudhry NA, Munir M, Hussain R, Tayab M. Eitiological Pattern of Septicemia at Three Hospitals in Lahore: JCPSP 2000; 10:375-9.
- 17 Mahmood A, Karamat KA, Butt T. Neonatal Sepsis: High Antibiotic Resistance of the Bacterial Pathogens in a Neonatal Intensive Care Unit in Karachi: JPMA. 2002; 52:348-50.
- 18 Viswanathan R, Singh AK, Basu S, Chatterjee S, Sardar S, Isaacs D; Multi-drug resistant gram negative bacilli causing early neonatal sepsis in India.; Arch Dis Child Fetal Neonatal Ed doi:10.1136/archdischild-2011-300097.
- 19 Karambin MM, Zarkesh M. Enterobacter, the Most Common Pathogen of Neonatal Septicemia in Rasht, Iran: Iran J Pediatr 2011; 21:83-7.
- 20 Ahmed A, Pervez MI, Paul BK, Bishwas KK. Clinical and Bacteriological Profile of Neonatal Septicaemia at A Community Level Medical College Hospital: JBCPS, 2011; 29:143-50.
- 21 Anwer SK, Mustafa S, Pariyani S, Ashraf S, Taufiq KM: Neonatal Sepsis: An Etiological Study: JPMA, 2000; 50:91-4.
- 22 Ojukwu JU, Abonyi LE, Ugwu J, Orji IK. Neonatal Septicaemia in High Risk Babies in South-Eastern Nigeria: J Perinat Med 2006; 34:166-72.

- 23 Anah MU, Udo JJ, Ochigbo SO, Abia-Bassey LN; Neonatal septicaemia in Calabar, Nigeria; Trop. Doct.; 2008; 38:126-8.
- 24 Bhutta ZA, Yousaf K; Neonatal Sepsis in Karachi: Factors Determining Outcome and Mortality; J Trop Pediatr 1997; 43:65-70.
- 25 Younas M, Rahim F; Early Neonatal Sepsis: An Etiological Study; JPMI 2011; 16:89-92.
- 26 Stoll BJ, Hansen NI, Sanchez PJ, Faix RG, Poindexter BB, Van Meurs KP, et al. Early Onset Neonatal Sepsis: The Burden of Group B Streptococcal and E.Coli Disease Continues; Pediatrics 2011; 127:817-26.
- 27 Darmstad GL, Maneesh B, Anita MZ. Oral Antibiotics in the Management of Serious Neonatal Bacterial Infections in Developing Country Communities; Pediatr Infect Dis J 2009; 28:31-6.
- 28 Ahmed MN, Khan NZ, Saha SK, Chowdhury KA, Muslima H, Law P, et al. Ciprofloxacin Treatment in Preterm Neonates in Bangladesh: Lack of Effects on Growth and Development; Pediatr Infect Dis J 2006; 25:1137-41.
- 29 Chaudhari S, Suryawanshi P, Ambardekar S, Chinchwadkar M, Kinare A. Safety profile of ciprofloxacin used for neonatal septicemia. Ind Pediatr 2004; 41:1246-51.
- 30 HabiburRasool C, Hassan AM, Habibullah M.; Neonatal Sepsis and use of Antibiotic in a Tertiary Care Hospital. Pak J Med Sci 2007; 3:78-81.



#### Statistics

Excerpts from the Uniform Requirements for Manuscripts Submitted to Biomedical Journals updated November 2003

Describe statistical methods with enough detail to enable a knowledgeable reader with access to the original data to verify the reported results. When possible, quantify findings and present them with appropriate indicators of uncertainty (such as confidence intervals). Avoid relying solely on statistical hypothesis testing. Such as the use of P values, which fails to convey important information about effect size. References for the design of the study and statistical methods should be to standard works when possible (with pages started). Define statistical terms, abbreviation, and most symbols. Specify the computer software used.

Sample references are available from www.icmje.org